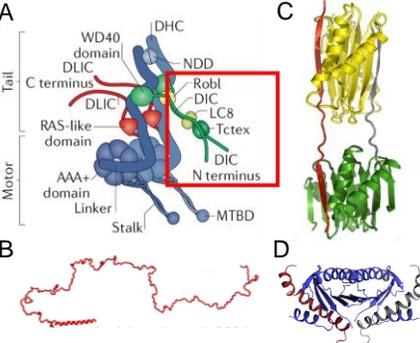


**Abstract**

Cytoplasmic dynein is a large, eukaryotic motor protein complex composed of heavy, intermediate, light intermediate, and light chain subunits. The motor domain of dynein has been well characterized, however in the tail domain there are almost 300 amino acids of disorder that remain missing from resolved dynein structures. This disorder lies in dynein's intermediate chain (IC), the key protein in dynein's cargo attachment subcomplex, involved in binding with dynein's light chain subunits as well as various non-dynein regulatory proteins. Two of dynein's regulatory proteins, dyactin's p150<sup>Glued</sup> subunit and a nuclear distribution protein (NudE) have both been shown to bind to the N-terminus, single  $\alpha$ -helix of IC in a variety of species. In this work, we characterize the binding interaction between IC and coiled-coiled segments of p150<sup>Glued</sup> and NudE in *Chaetomium thermophilum* (CT), a thermophilic filamentous fungus and a novel choice for dynein studies. With the use of isothermal titration calorimetry (ITC), analytical ultracentrifugation (AUC), and nuclear magnetic resonance (NMR) we have explored the importance of the level of disorder in a transient, secondary helix (H2) in IC's binding affinity to regulatory proteins. Furthermore, by studying both a small, 88 residue construct of IC, as well as a much larger 260 residue construct, we have demonstrated that multivalency and disorder underlie the regulation of the IC/p150<sup>Glued</sup> and IC/NudE interactions.

**Background**

**Dimeric dynein light chains bring together two disordered, monomeric chains of N-IC to create a subcomplex scaffold**



(A) Much of dynein is structurally well characterized<sup>1</sup>. The cargo attachment subcomplex however, (boxed in red) has much more ambiguity due to its disorder and flexibility. (B) The N-term of IC is an almost completely disordered monomeric chain. (C-D) Solved crystal structures of Tctex and LC8 (C) and LC7 (D) in complex with IC. The dimeric light chains bring together two chains of IC to form a polybivalent scaffold. IC adopts light chain structure at the binding sites, while linkers remain disordered<sup>2-10</sup>.

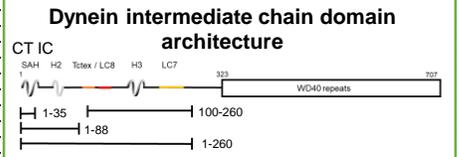


**Role of Disorder in the Regulation of Dynein Intermediate Chain**

Kayla A. Jara<sup>1</sup>, Nikolaus M. Loening<sup>2</sup>, and Elisar J. Barbar<sup>1</sup>

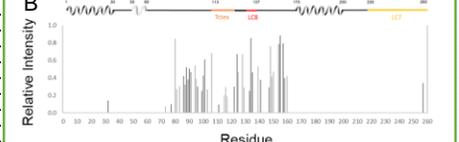
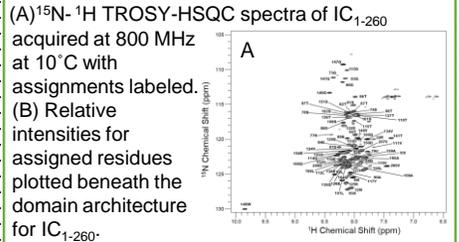
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**N-IC is disordered**

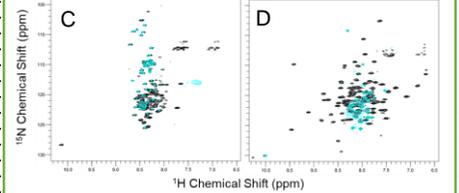


IC from *Chaetomium thermophilum* (CT IC) is predicted to have an N-terminal single  $\alpha$ -helix (SAH), a transient/nascent second helix (H2), and a more C-terminal helix (H3). Based on sequence motifs, the Tctex (orange), LC8 (red), and LC7 (yellow) binding sites are predicted to be in regions similar to Drosophila IC. The C-terminal region is predicted to contain seven WD40 repeat domains. IC<sub>1-88</sub>, IC<sub>1-260</sub>, IC<sub>100-260</sub>, and IC<sub>140-260</sub> show the constructs used in this work. IC<sub>1-35</sub> is a construct used in prior work.

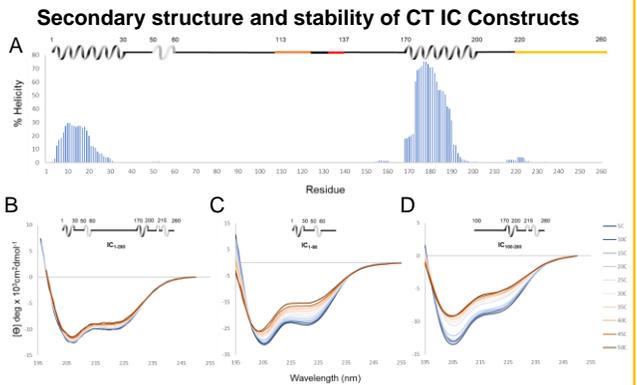
**Characterization of disorder in CT IC<sub>1-260</sub> by NMR**



(C-D) Overlays of a CLEANEX experiment (light blue) onto the <sup>15</sup>N-<sup>1</sup>H TROSY-HSQC spectrum (black) for both 10°C (C) and 40°C (D).

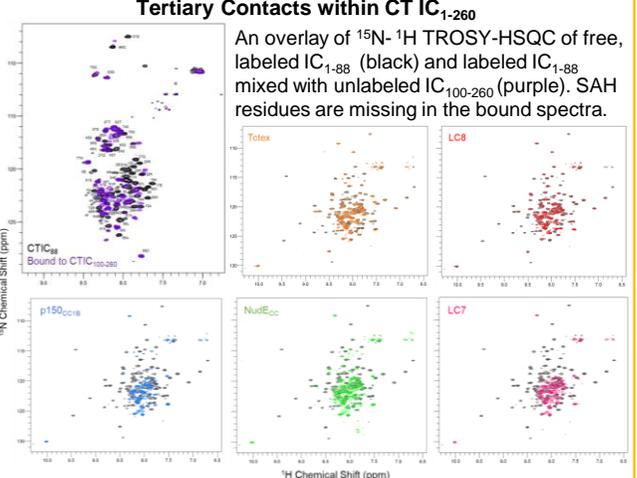


**IC<sub>1-260</sub> is stabilized by a H3**



(A) Agadir helix prediction of IC<sub>1-260</sub>, plotting %Helicity by residue. Temperature-dependent CD spectra of (B) IC<sub>1-260</sub>, (C) IC<sub>1-88</sub>, and (D) IC<sub>100-260</sub>. Spectral shape for all constructs indicates a mixture of  $\alpha$ -helical secondary structure as well as regions of intrinsic disorder. Loss in structure, or lack thereof, over a temperature range of 5-50°C indicate how each construct varies in stability.

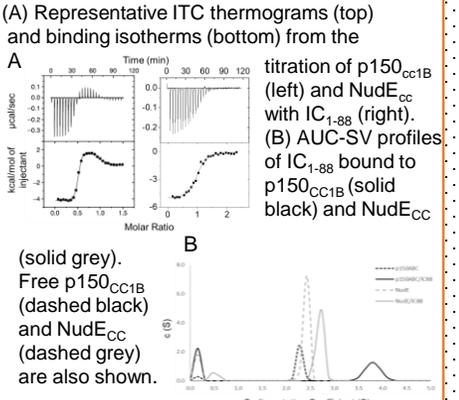
**Long-range interactions between IC<sub>1-260</sub> Helices**



Overlays of free IC<sub>1-260</sub> (black) and binding partners at 40°C. Binding patterns of partners bound at the N-term (p150 & NudE) match that of LC7 bound at the C-term. Binding at sights more central linker regions (Tctex & LC8) show their own distinct pattern.

**IC/p150 & IC/NudE Interactions**

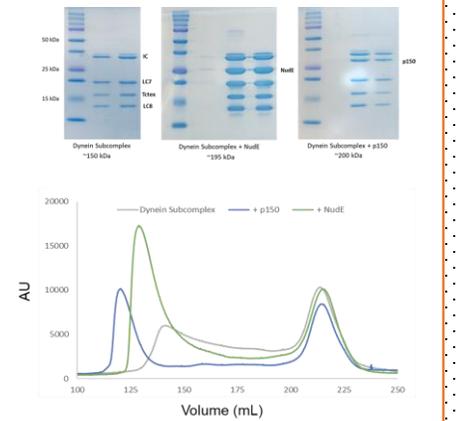
**Binding interactions of CT IC<sub>1-88</sub> with p150<sub>CC1B</sub> and NudE<sub>CC</sub>**



(A) Representative ITC thermograms (top) and binding isotherms (bottom) from the titration of p150<sub>CC1B</sub> (left) and NudE<sub>CC</sub> (right) with IC<sub>1-88</sub> (right). (B) AUC-SV profiles of IC<sub>1-88</sub> bound to p150<sub>CC1B</sub> (solid black) and NudE<sub>CC</sub> (dashed grey) are also shown.

**Subcomplex Reconstitution**

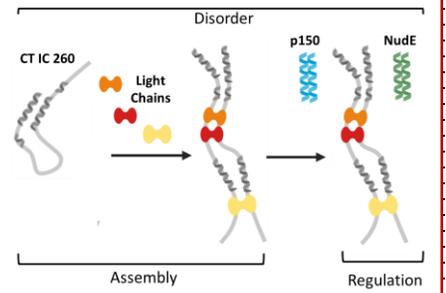
**Purification and characterization of reconstituted dynein subcomplexes**



SEC traces of the dynein subcomplex (IC/light chains) (grey) and the dynein subcomplex with p150<sub>CC1B</sub> (blue) or NudE<sub>CC</sub> (green). SDS-PAGE gels of fractions collected from SEC for all complexes showing all expected proteins

**Conclusions**

**Disorder, assembly, and regulation in the dynein attachment subcomplex**



From our studies with CT IC<sub>1-260</sub> we conclude:

- CT IC<sub>1-260</sub> is largely disordered with regions of helicity that impart long-range tertiary contacts between the N and C-terminus
- Tertiary contacts cause free CT IC<sub>1-260</sub> to adopt a more compact structure that is stabilizing compared to smaller constructs
- This construct of IC allows for study of complex binding interactions, including the successful reconstitution of various forms of fully bound subcomplexes.
- Binding of the three dynein light chains cause CT IC<sub>1-260</sub> to form an extended polybivalent scaffold and bind to p150<sub>CC1B</sub> and NudE<sub>CC</sub> with greater affinity than free CT IC<sub>1-260</sub> alone.
- Disorder and assembly of the cargo attachment subcomplex are critical to its regulation

**Acknowledgements**

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